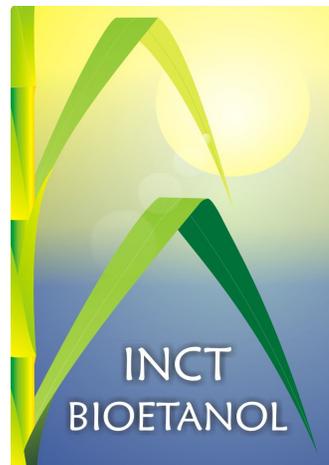


Carbon pathways in sugarcane and miscanthus : *targets to improve them artificially*

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& Amanda Pereira de Souza

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Feedstock biochemistry applied to Biofuels, Fulbright, 2009

The wall in the context of plant composition

Obtained from CO₂ and water

| | | |
|-----|---|-----------------|
| 96% | { | Carbon.....45% |
| | | Oxygen.....45% |
| | | Hydrogen.....6% |

**Cellulose,
hemicelluloses & pectins
96-10%=86%**

Macronutrients

| | | |
|------|---|--|
| 3.6% | { | Nitrogen.....1.5% X 6.25 = 9.4% (10%) |
| | | Potassium.....1.0% X |
| | | Calcium.....0.5% |
| | | Magnesium.....0.2% |
| | | Phosphorous.....0.2% X |
| | | Sulfur.....0.1% X |
| | | Silicium.....0.1% |

**Proteins and
Nucleic acids**

Pectins = 0.7%

Pectins? = 0.7%

Micronutrients

| | | |
|------|---|---------------------------|
| 0.4% | { | Boron..... |
| | | Manganese..... |
| | | Chloride..... X |
| | | Iron..... X |
| | | Sodium..... X |
| | | Zinc..... X |
| | | Copper..... X |
| | | Nickel..... X |
| | | Molibdenium..... X |

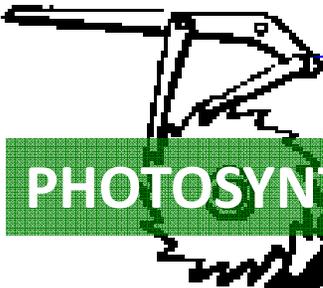
Pectins = traces

Lipids are approximately 15% of plant tissues

**Thus, the wall corresponds
to ca. 70 % of the plant**

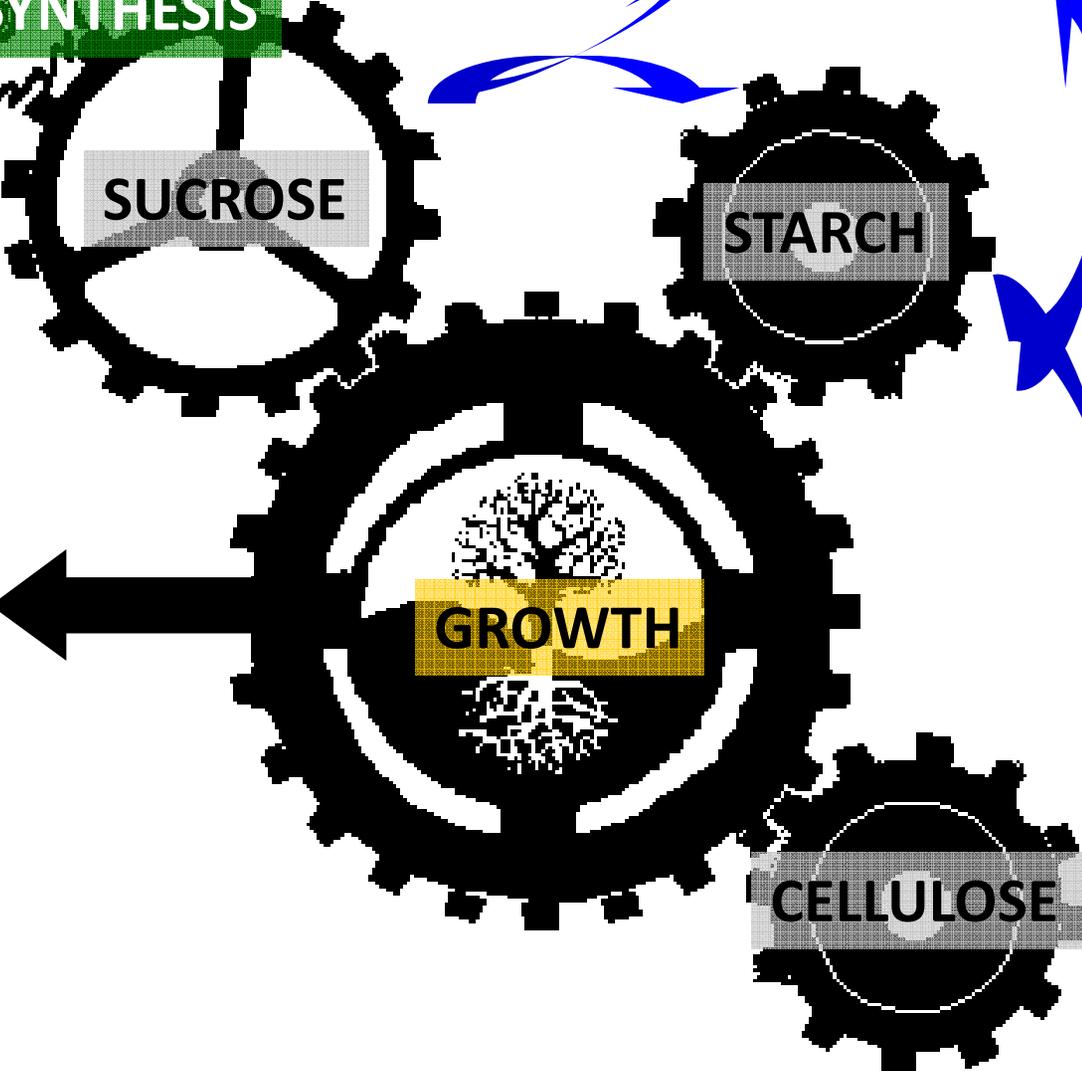
*In sugarne = leaves contain 68% and
stem 50% plus 18% of sucrose*

Light , water and nutrients



PHOTOSYNTHESIS

CO₂

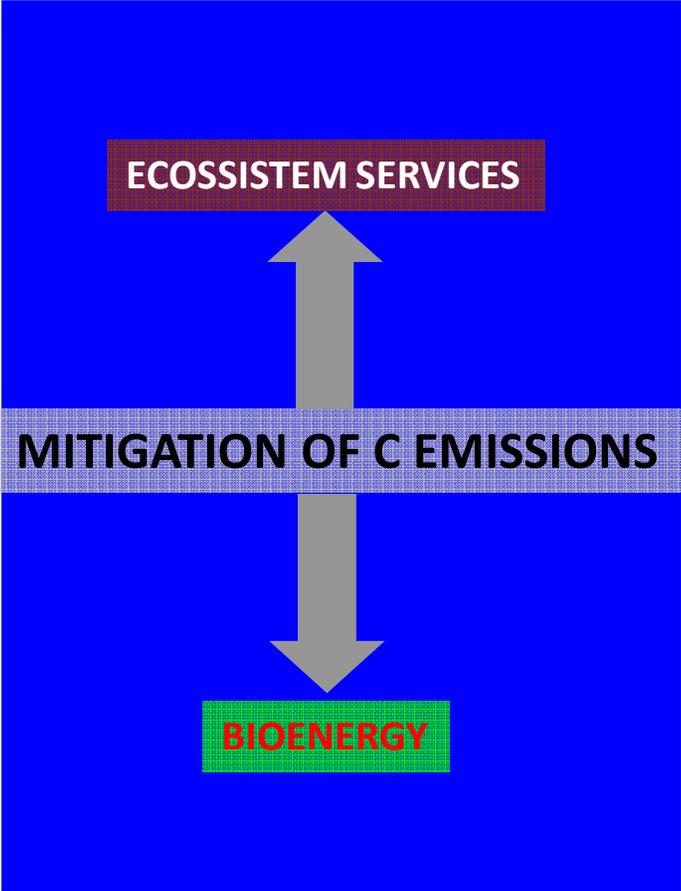


SUCROSE

STARCH

GROWTH

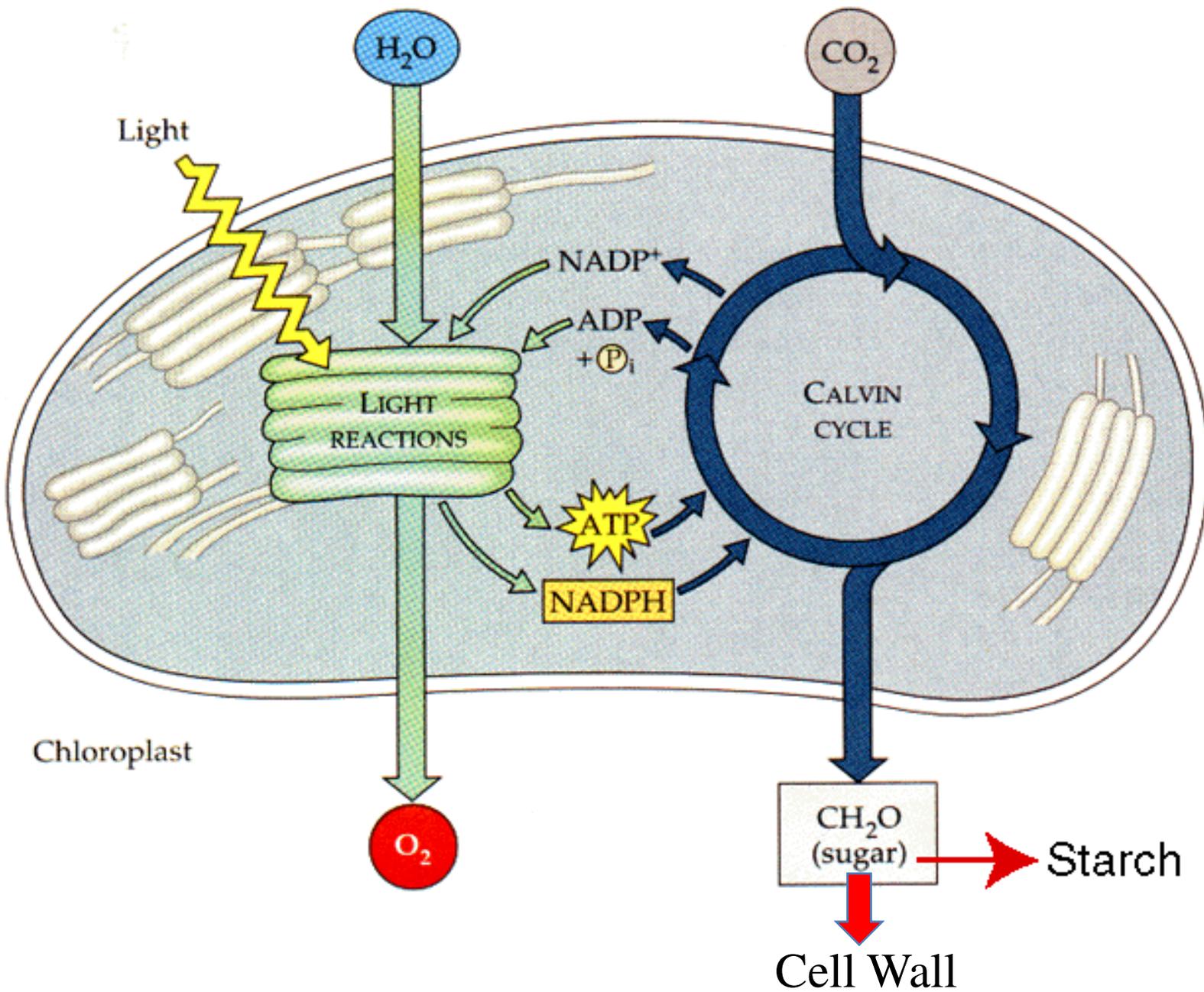
CELLULOSE

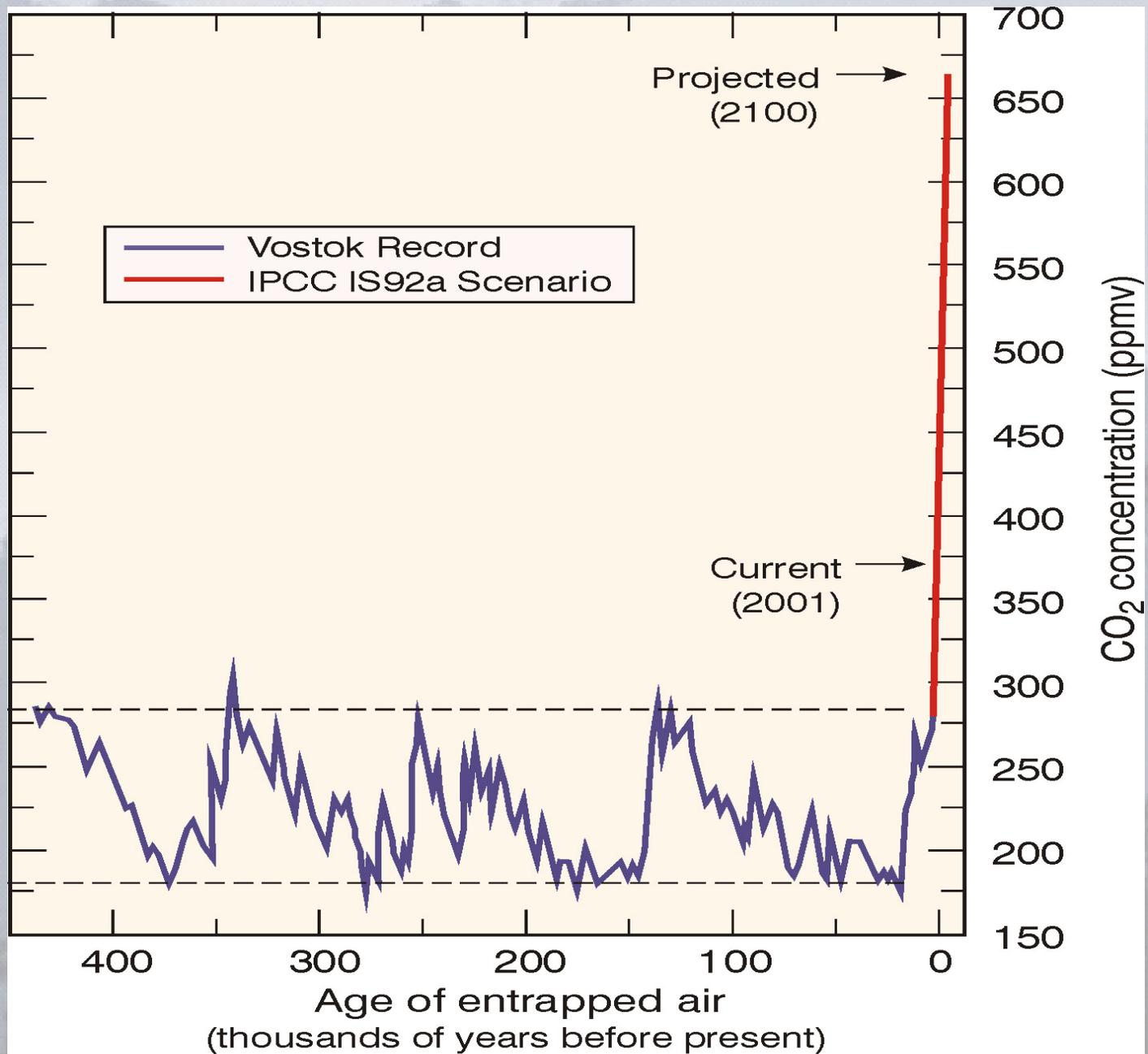


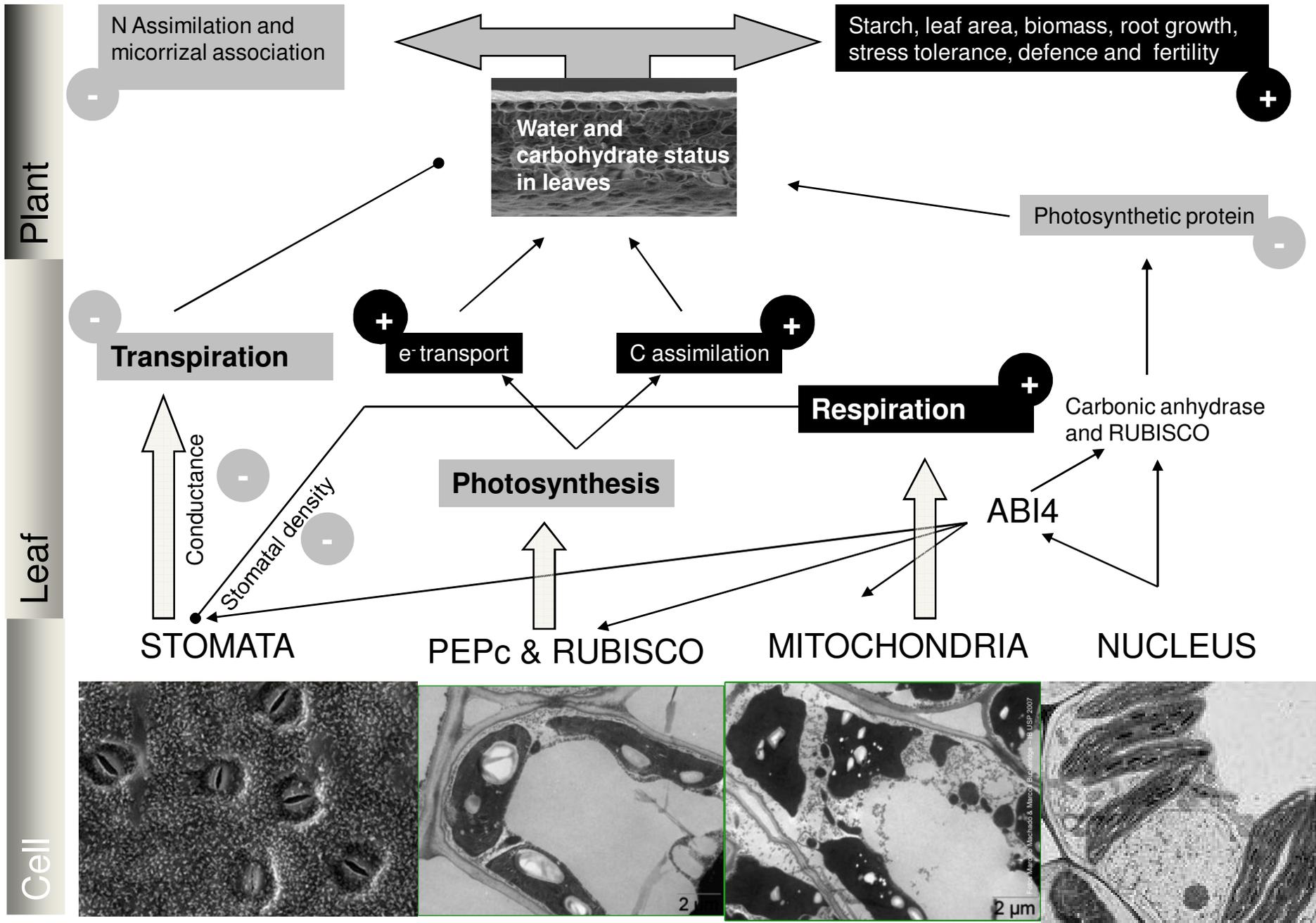
ECOSYSTEM SERVICES

MITIGATION OF C EMISSIONS

BIOENERGY





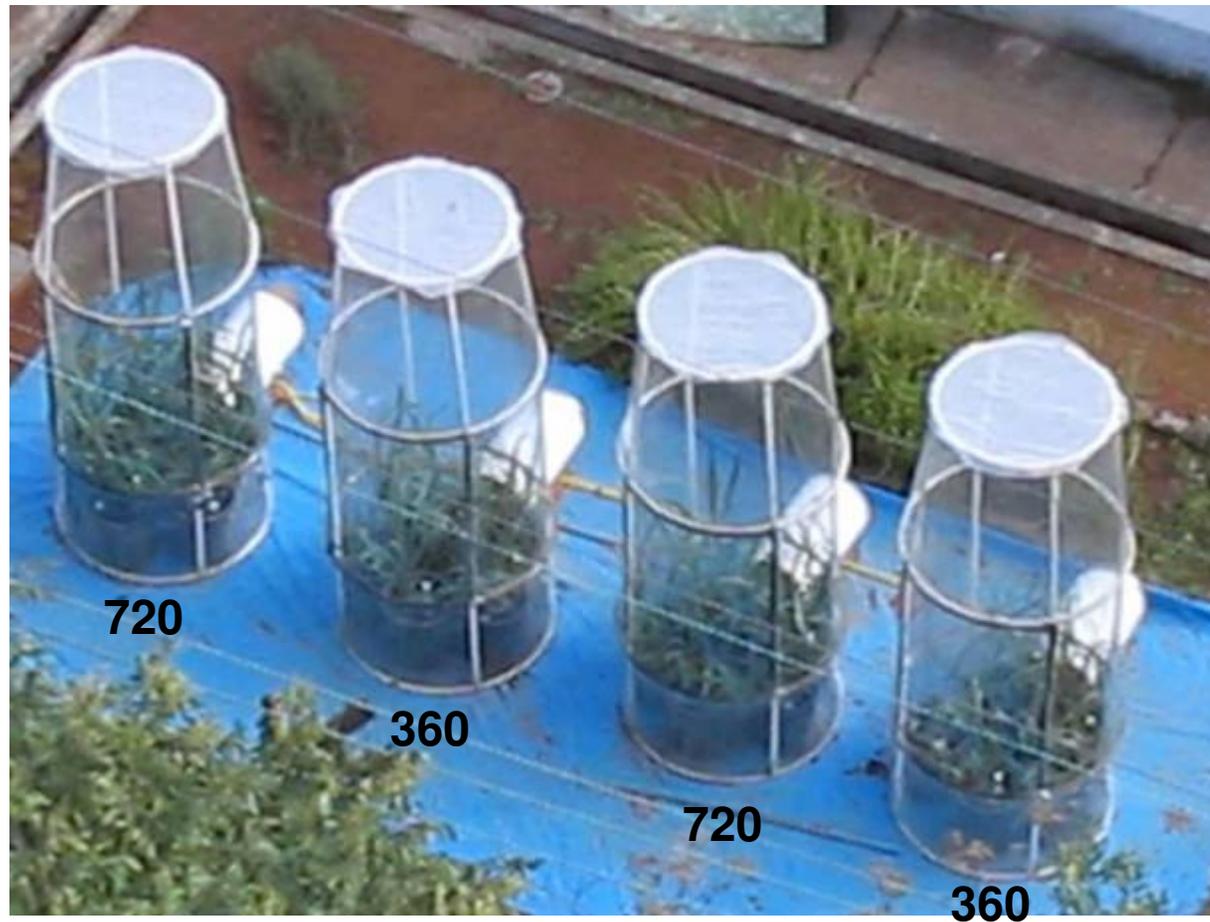


Elevated CO₂ increases photosynthesis, biomass and productivity, and modifies gene expression in sugarcane

AMANDA PEREIRA DE SOUZA¹, MARILIA GASPAR², EMERSON ALVES DA SILVA², EUGÊNIO CÉSAR ULIAN², ALESSANDRO JAQUIEL WACLAWOVSKY³, MILTON YUTAKA NISHIYAMA JR.³, RENATO VICENTINI DOS SANTOS⁵, MARCELO MENOSSI TEIXEIRA⁵, GLAUCIA MENDES SOUZA² & MARCOS SILVEIRA BUCKERIDGE¹

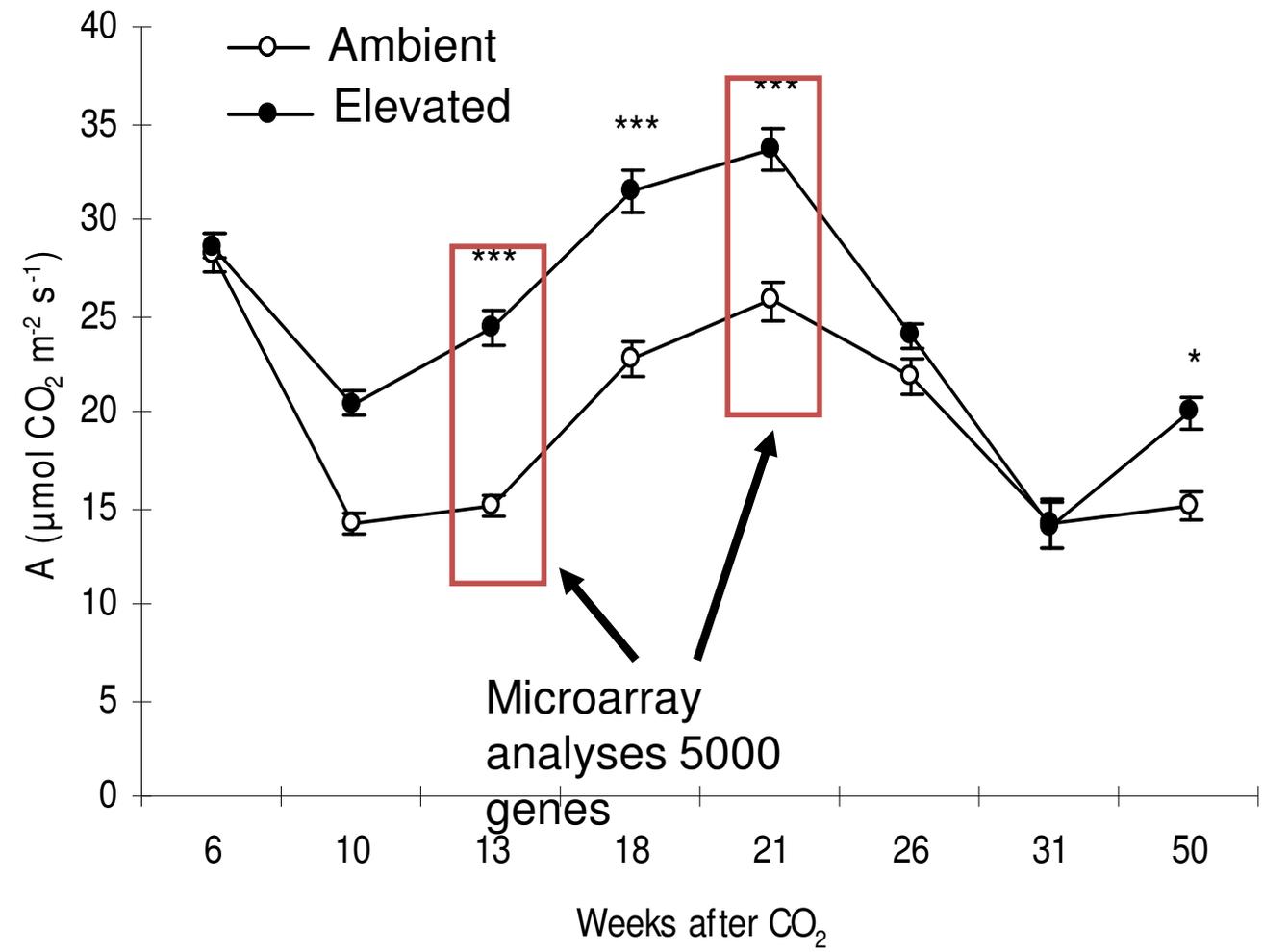
¹*Departamento de Botânica, Instituto de Biociências and* ²*Departamento de Bioquímica, Instituto de Química, Universidade de São Paulo, São Paulo, Brazil,* ³*Seção de Fisiologia e Bioquímica de Plantas, Instituto de Botânica, São Paulo, Brazil,* ⁴*Centro de Tecnologia Canavieira, Piracicaba, Brazil and* ⁵*Centro de Biologia Molecular e Engenharia Genética, Universidade Estadual de Campinas, Campinas, São Paulo, Brazil*

Sugar cane in the open top chambers 2005



Funded by Centro de Tecnologia Canavieira - Piracicaba

Photosynthesis in sugarcane growing under elevated CO₂



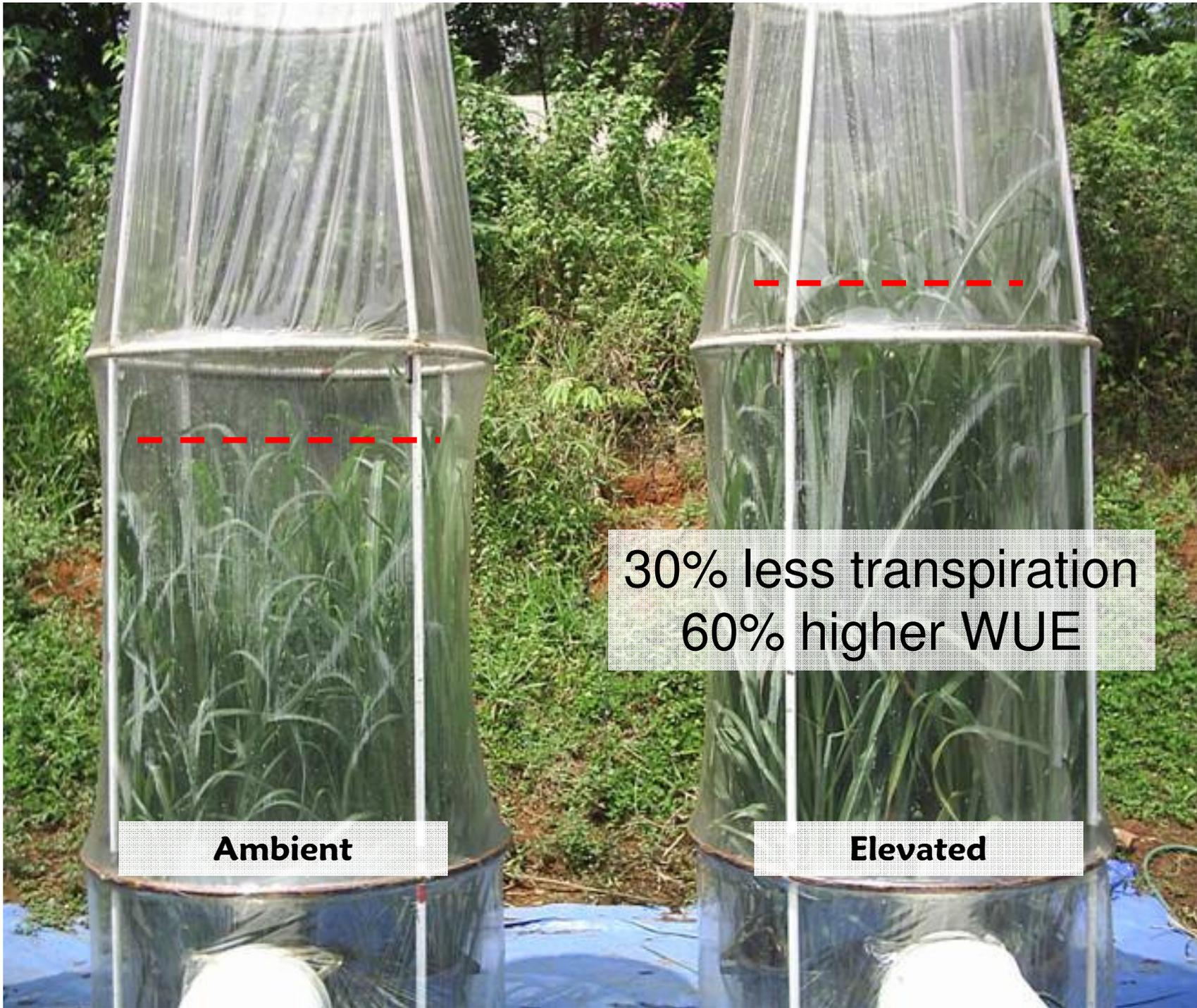
Microarray analysis of the CO₂ experiments

3 months

| Categories | Gene description | Ratio (elevated/ambient) |
|--|---|-------------------------------------|
| Development | light-induced protein | 1,194 |
| Photosynthesis | photosystem II protein K; psbK | 1,315 |
| Photosynthesis | Ferredoxin I; chloroplast precursor | 1,26 |
| Photosynthesis | photosystem I reaction centre subunit n, chloroplast precursor | 1,583 |
| Cell wall metabolism | xyloglucan endo- transglycosylase/hydrolase | 2,582 |
| Photosynthesis | Chlorophyll A-B binding protein | 1,508 |
| Stress response | ASR-like | 1,735 |
| Lipid, fatty-acid and isoprenoid metabolism | AE9 stearyl-ACP desaturase | 3,59 |
| Carbohydrate metabolism | beta-glucosidase isozyme 2 precursor | -2,189 |
| Carbohydrate metabolism | putative glucose-6-phosphate dehydrogenase | -1,232 |
| Protein metabolism | translational initiation factor eIF-4A | -1,606 |

5 months

| | | |
|--------------------------------------|--|--------|
| Protein metabolism | large ribosomal protein 2 | 1,454 |
| Carbohydr. metabolism/Photosynthesis | phosphoenolpyruvate carboxylase | 1,245 |
| Cell wall metabolism | Alpha-L-arabinofuranosidase | 1,37 |
| Protein metabolism | cathepsin B-like cysteine protease | 1,349 |
| Development | dormancy-associated protein | 2,299 |
| Transporters | Sugar transporter | 1,252 |
| Receptors | serine/threonine-protein kinase NAK | 1,706 |
| | unknow | 1,271 |
| Protein metabolism | putative glutamate-tRNA ligase | 1,504 |
| Protein metabolism | cathepsin B-like cysteine protease | 1,395 |
| Protein metabolism | Aldo/keto reductase; Sigma-54 factor | 1,229 |
| | putative nucleostemin (GTPase of unknown function) | 1,397 |
| Development | putative auxin-independent growth promoter | 1,909 |
| Transcription | pre-mRNA splicing factor | 1,541 |
| Nucleic acid metabolism | chromodomain-helicase-DNA-binding protein | 0,17 |
| Stress response | dehydrin | -1,42 |
| | unknow | -1,398 |
| Secondary metabolism | caffeoyl-CoA 3-O-methyltransferase 1 | -1,354 |
| Carbohydrate metabolism | cell wall invertase | -1,615 |
| Stress response | ferritin | -2,768 |
| Protein metabolism | C2 domain-containing protein-like | -1,342 |
| Pathogenicity | Thaumatococcus | -1,367 |
| Transcription | auxin response factor 2 | -1,352 |
| Development | Lateral organ boundaries protein | -1,494 |
| Cell cycle | kelch repeat-containing F-box family protein | -1,541 |
| Cell cycle | cyclin H-1 | -1,632 |



Ambient

Elevated

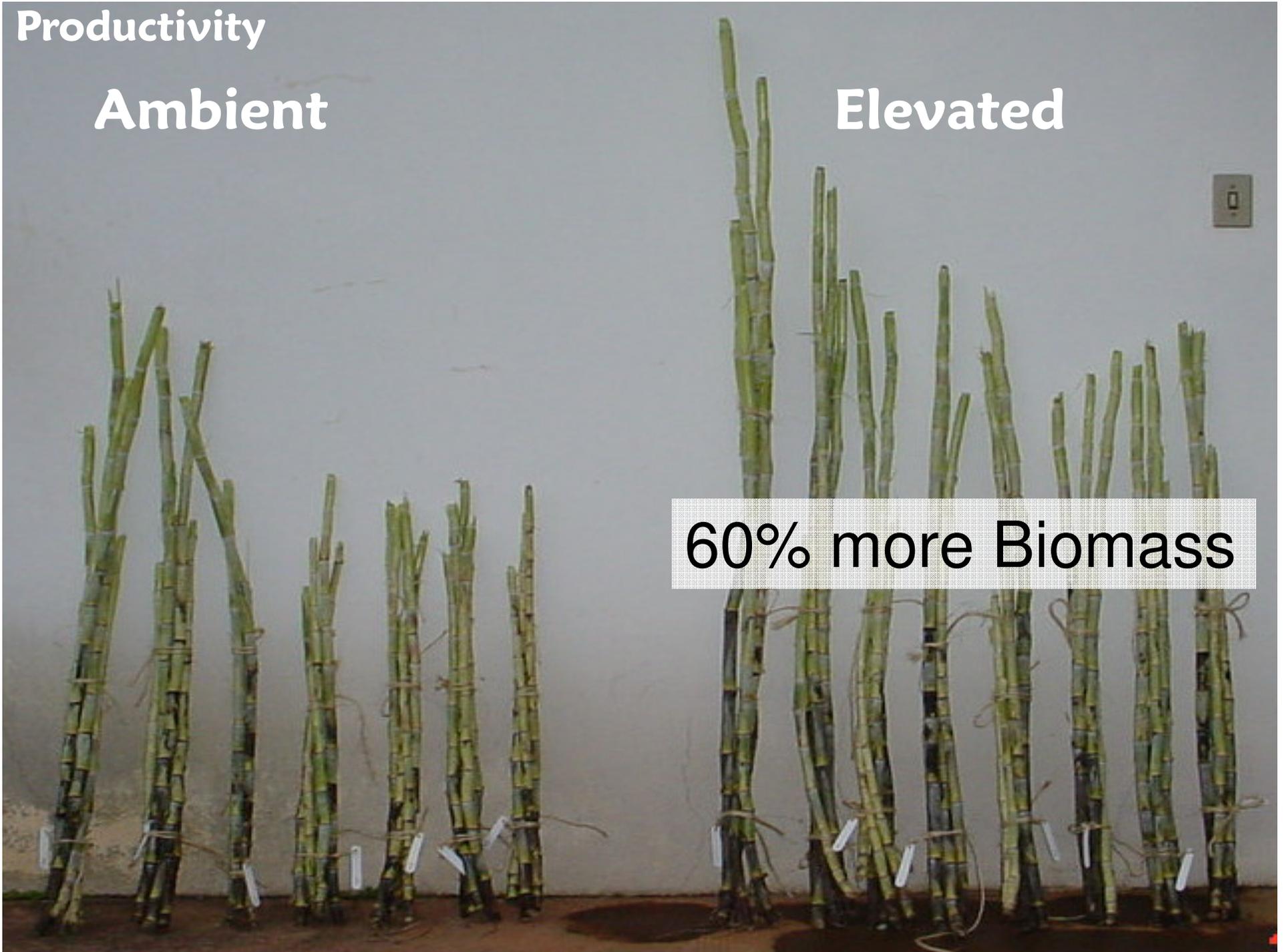
30% less transpiration
60% higher WUE

Productivity

Ambient

Elevated

60% more Biomass



SUGARS AND FIBERS

| | BRIX | Fiber(% FW) | Sucrose (% FW) |
|-----------------|-----------------|--------------------|-----------------------|
| Ambient | 7.17 ± 0.21 | 6.62 ± 0.13 | 2.18 ± 0.20 |
| Elevated | 7.75 ± 0.17 | 7.13 ± 0.21 | $2.82 \pm 0.14^*$ |

Speculative calculations

2087 – 653 millions of tons
Etanol – 20 millions of m³

2050 – 960 millions of tons
Etanol - 32 millions of m³

However, there are projections for much higher production by 2017

GENES ARE UP, BUT ARE THERE
COMPATIBLE BIOCHEMICAL AND
PHYSIOLOGICAL RESPONSES?

INCT
BIOETANOL

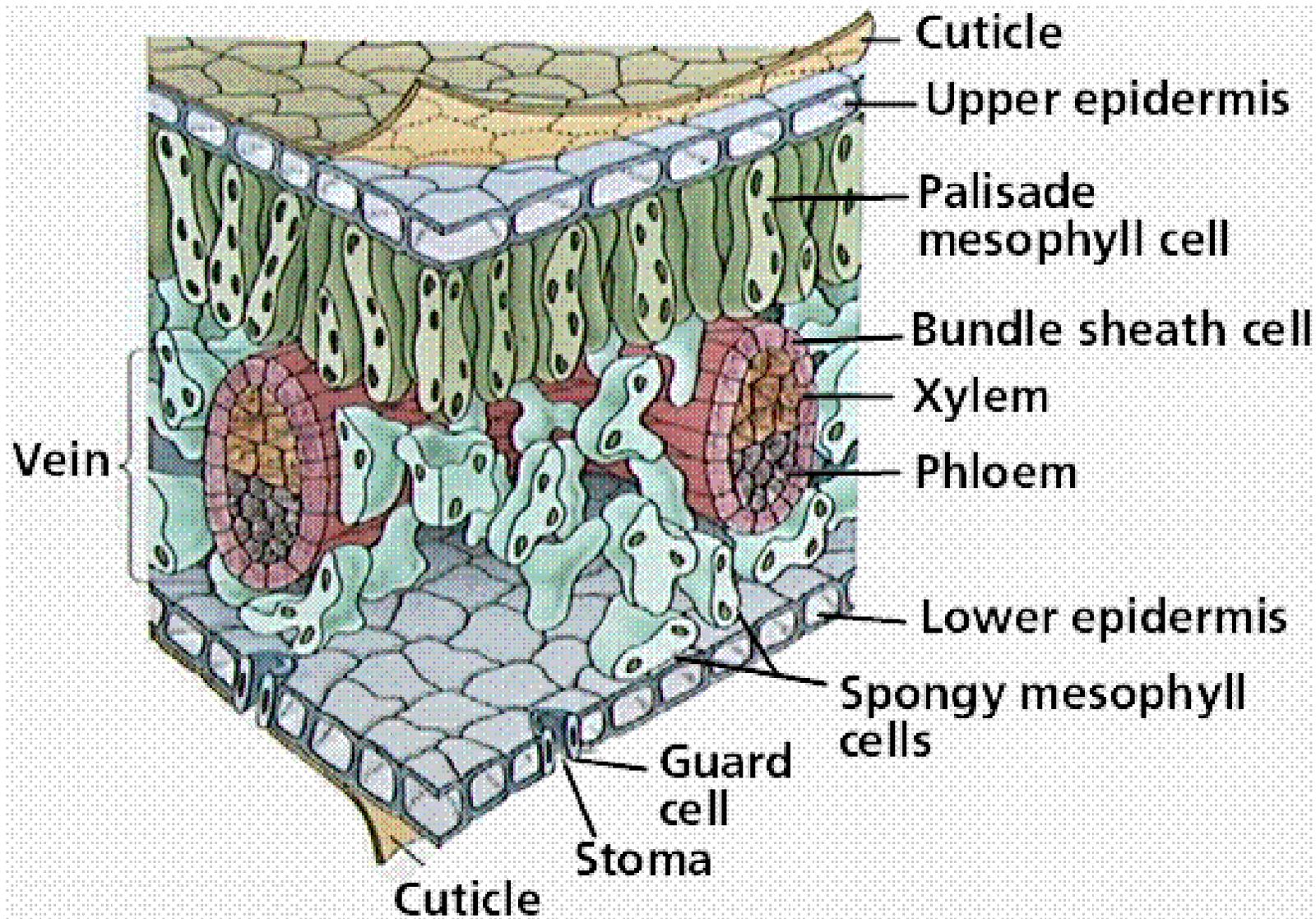
<http://bioethanolbrazil.wordpress.com>

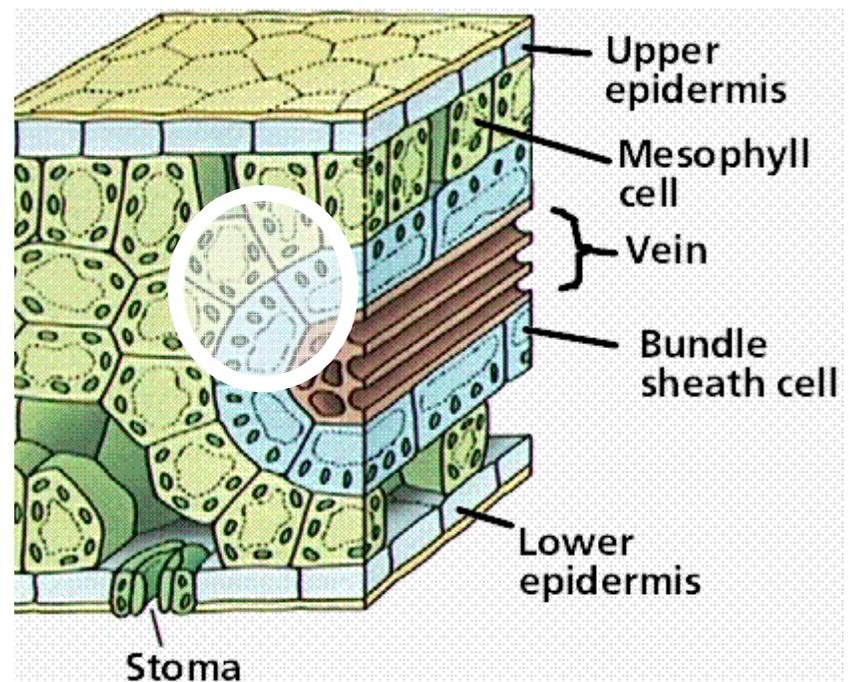
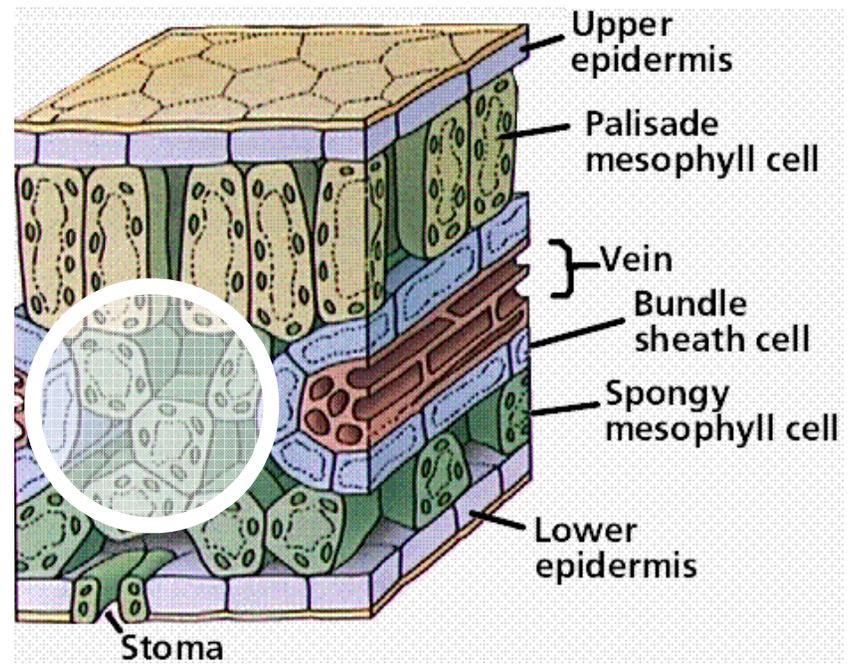
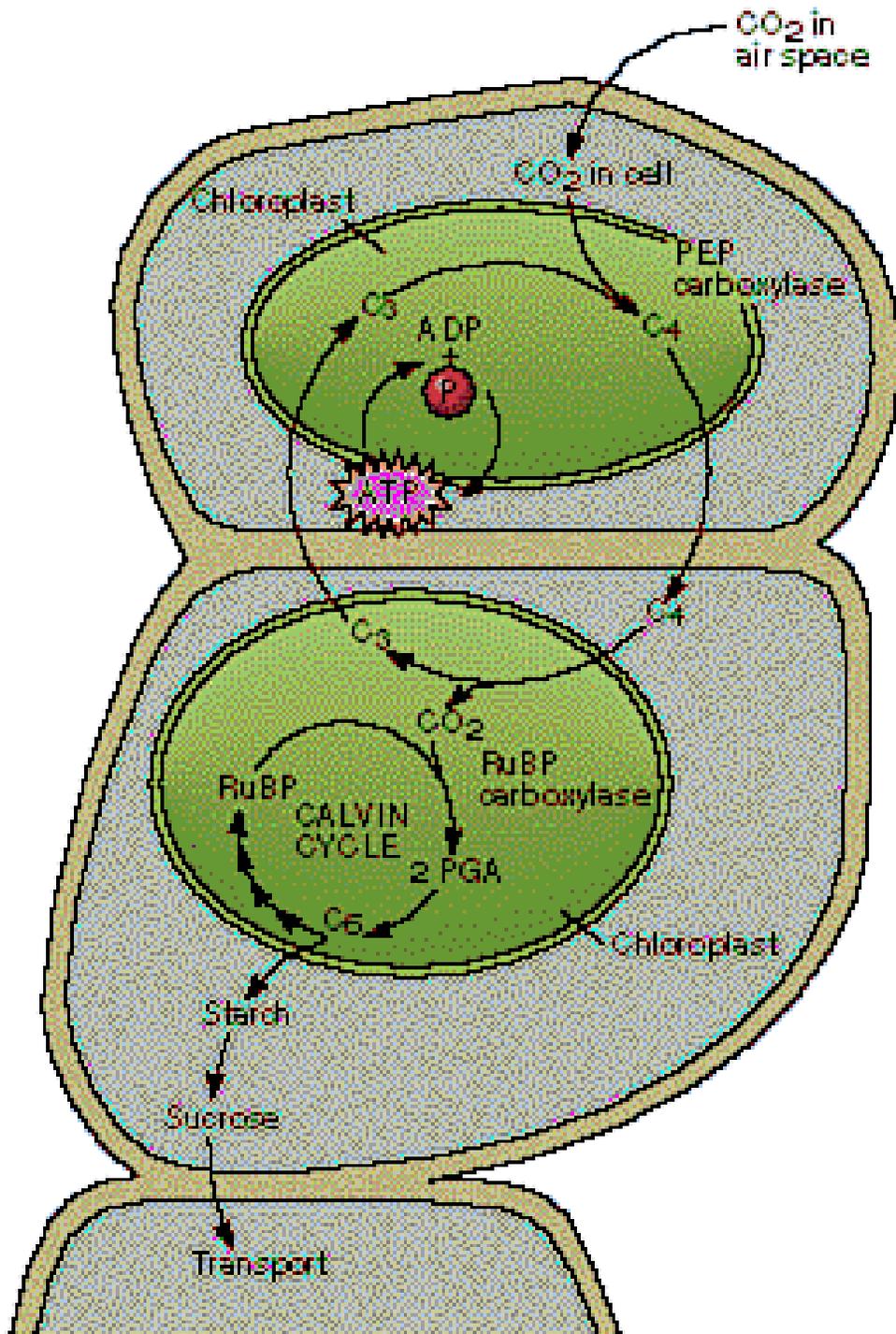




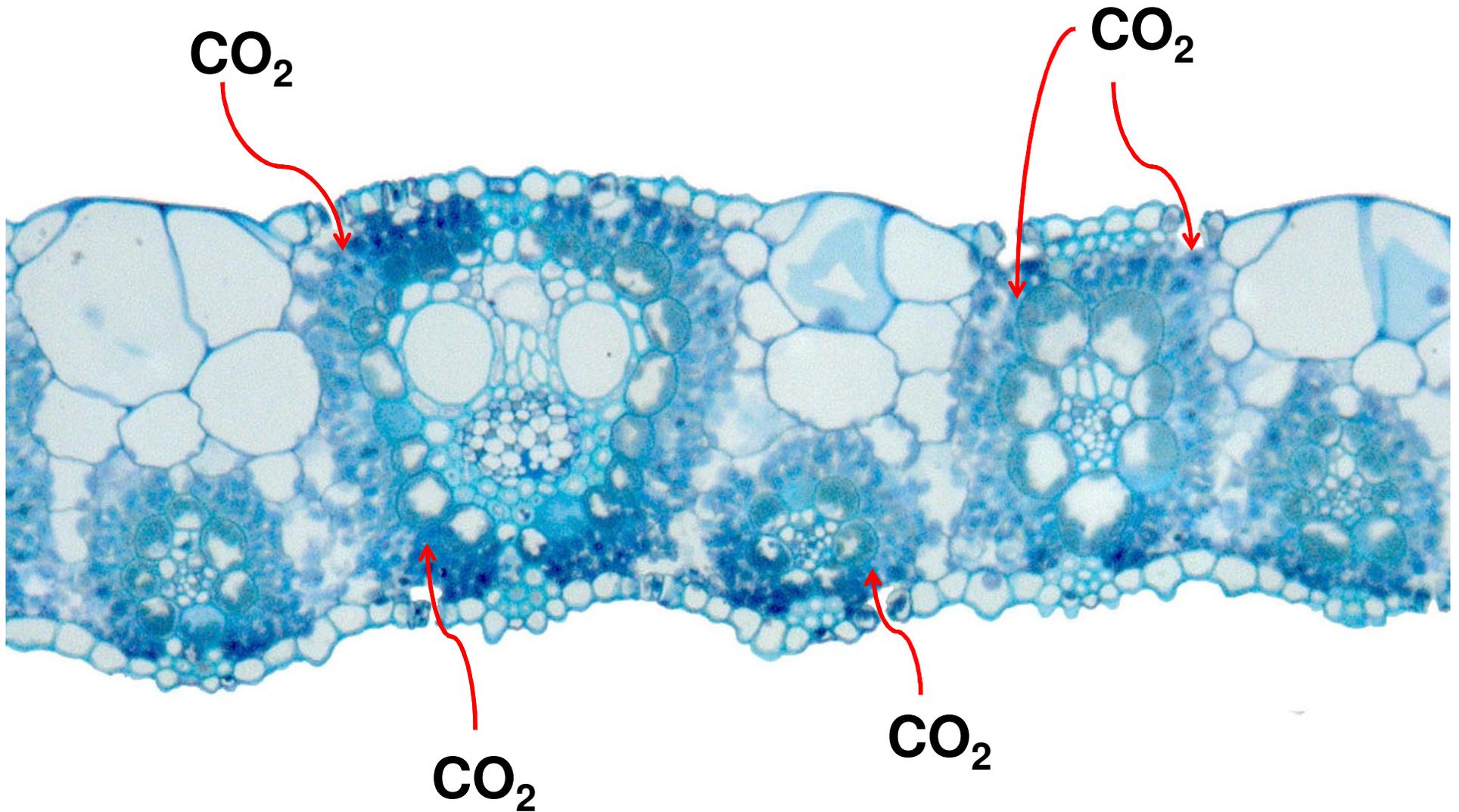
2008

75 dias em elevado CO₂



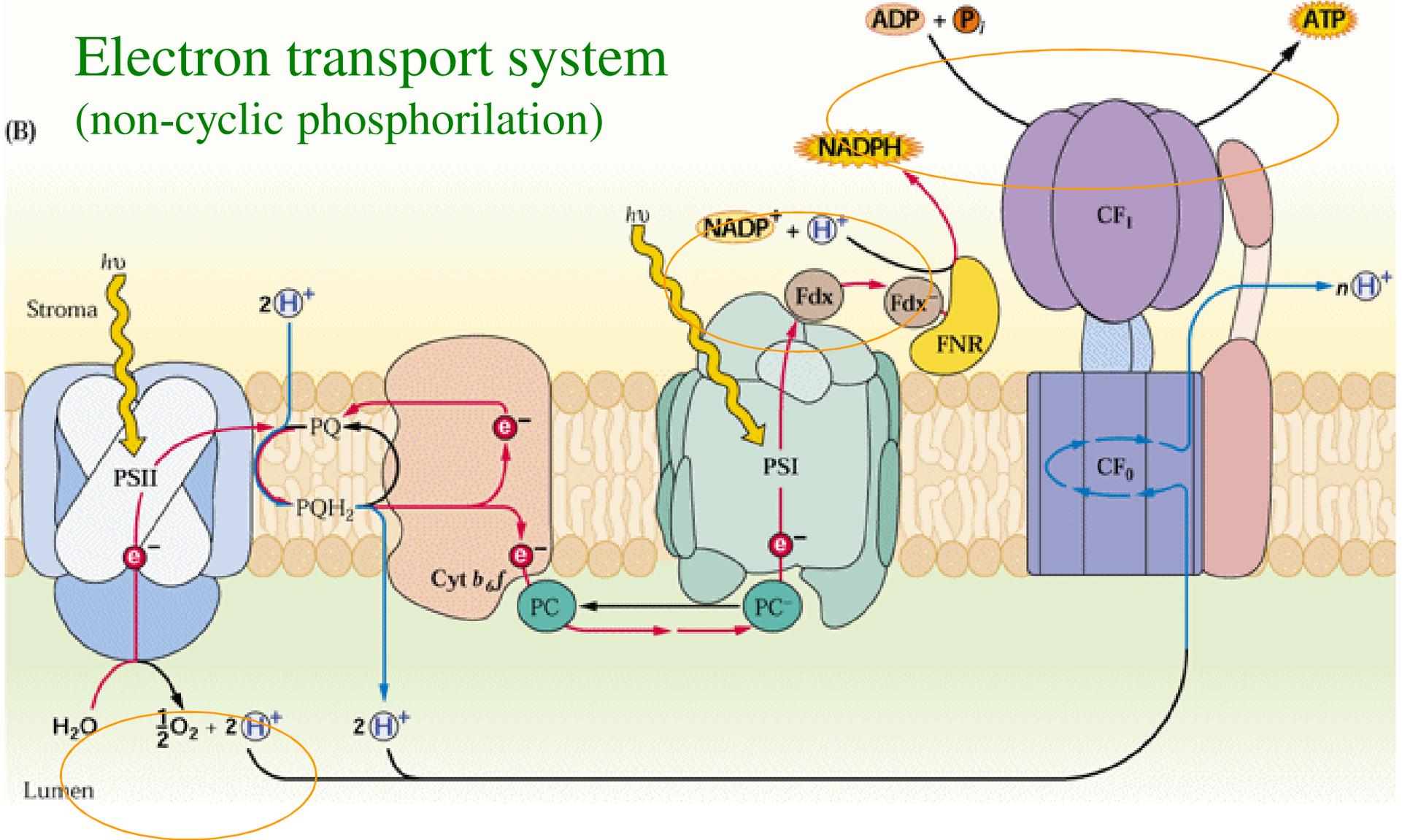


Sugarcane leaves:
CO₂ enters at both sides



Electron transport system

(B) (non-cyclic phosphorilation)





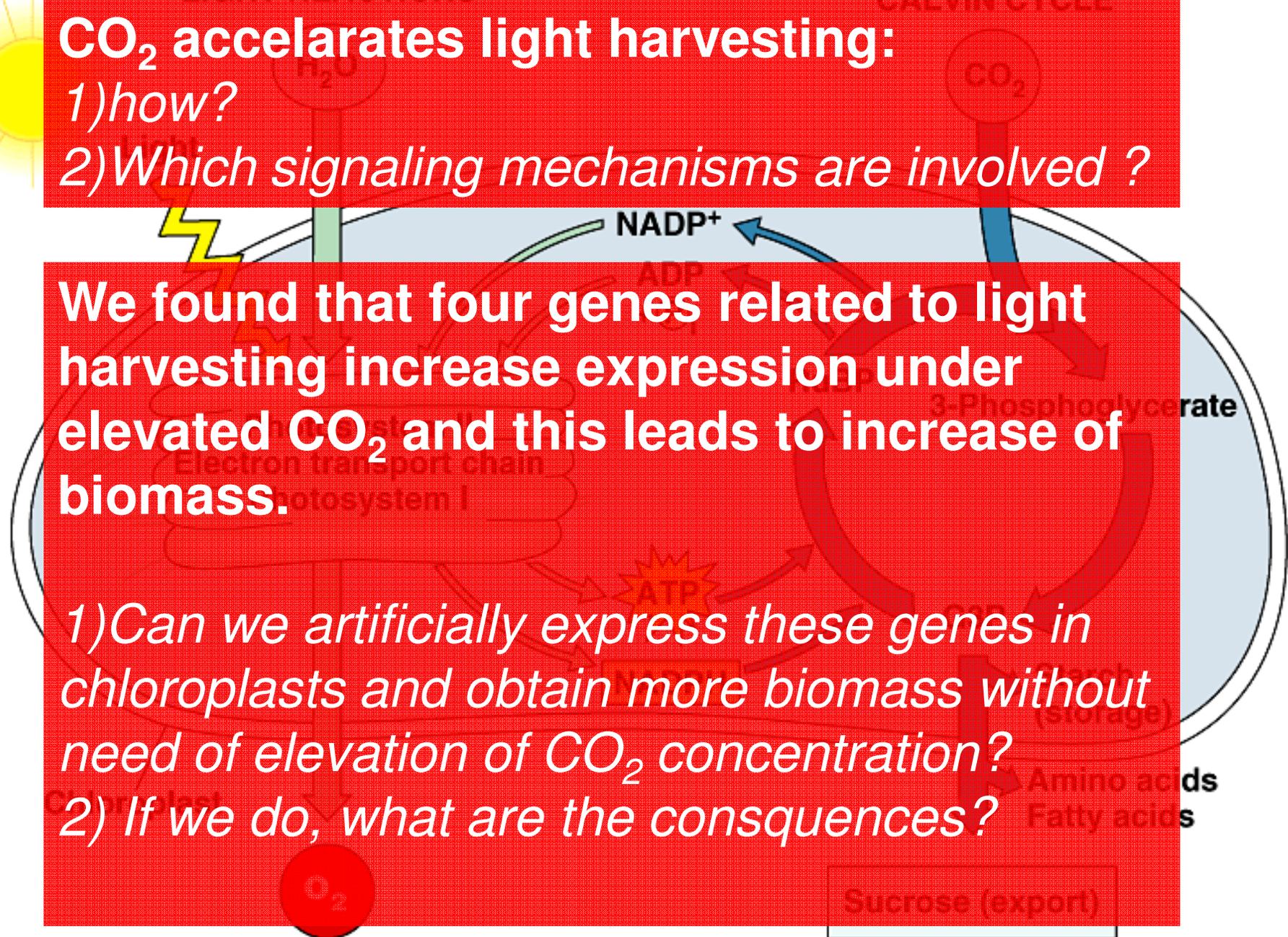
LIGHT REACTIONS

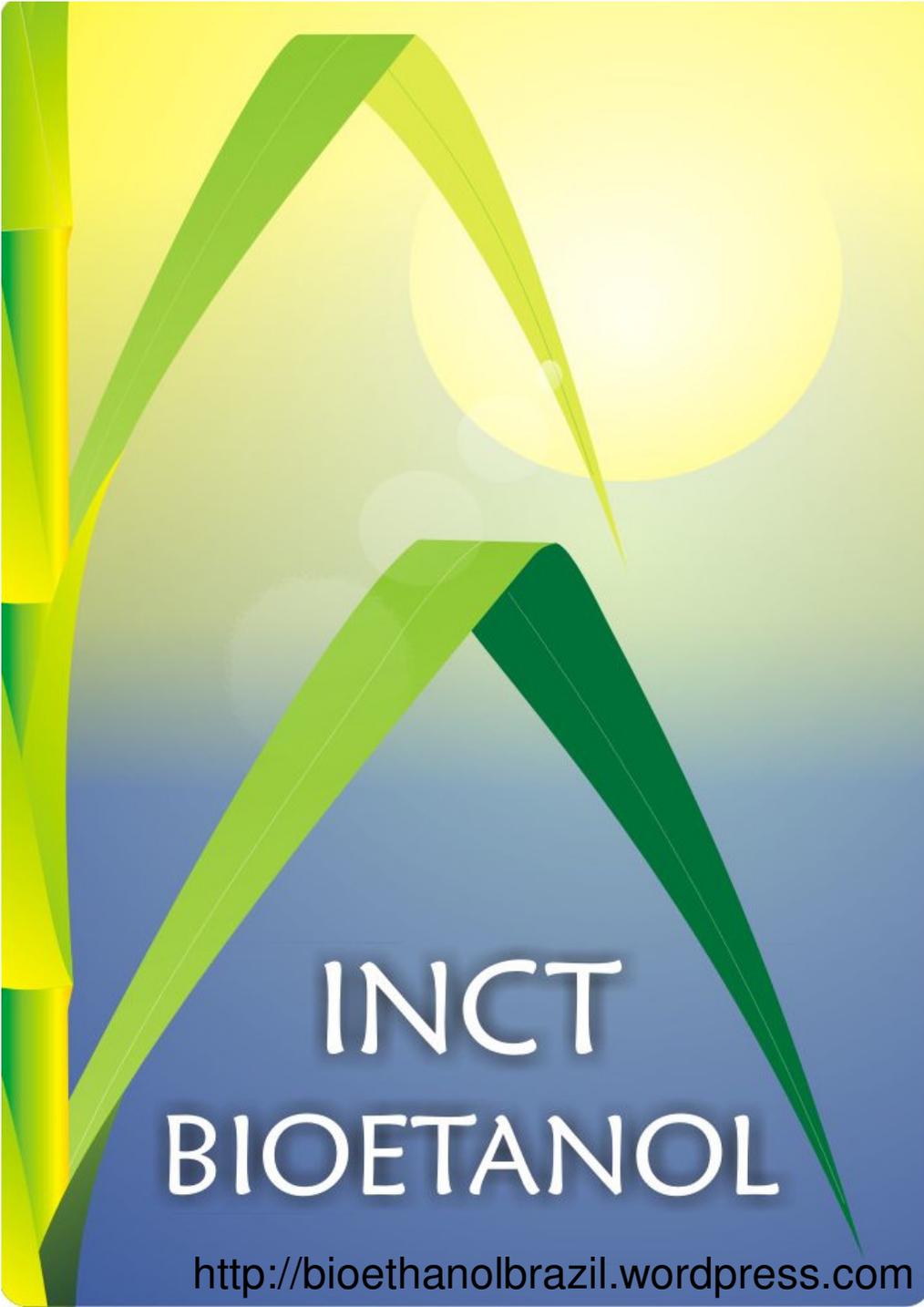
CALVIN CYCLE

CO₂ accelerates light harvesting:
1) how?
2) Which signaling mechanisms are involved ?

We found that four genes related to light harvesting increase expression under elevated CO₂ and this leads to increase of biomass.

1) Can we artificially express these genes in chloroplasts and obtain more biomass without need of elevation of CO₂ concentration?
2) If we do, what are the consequences?





INCT
BIOETANOL

<http://bioethanolbrazil.wordpress.com>

E METABOLISM IN SUGARCANE



Sugarcane stores sucrose and is hardly capable to produce starch

STORAGE



CO₂

Photosynthesis

Triose-P

Hexose-P

Glc & Fru

Sucrose

ADP-GPP

ADP-Glc

Starch

UDP-GPP

HK

INV

SPP

Sucrose-P

SPS

UDP-Glc

Cell Wall

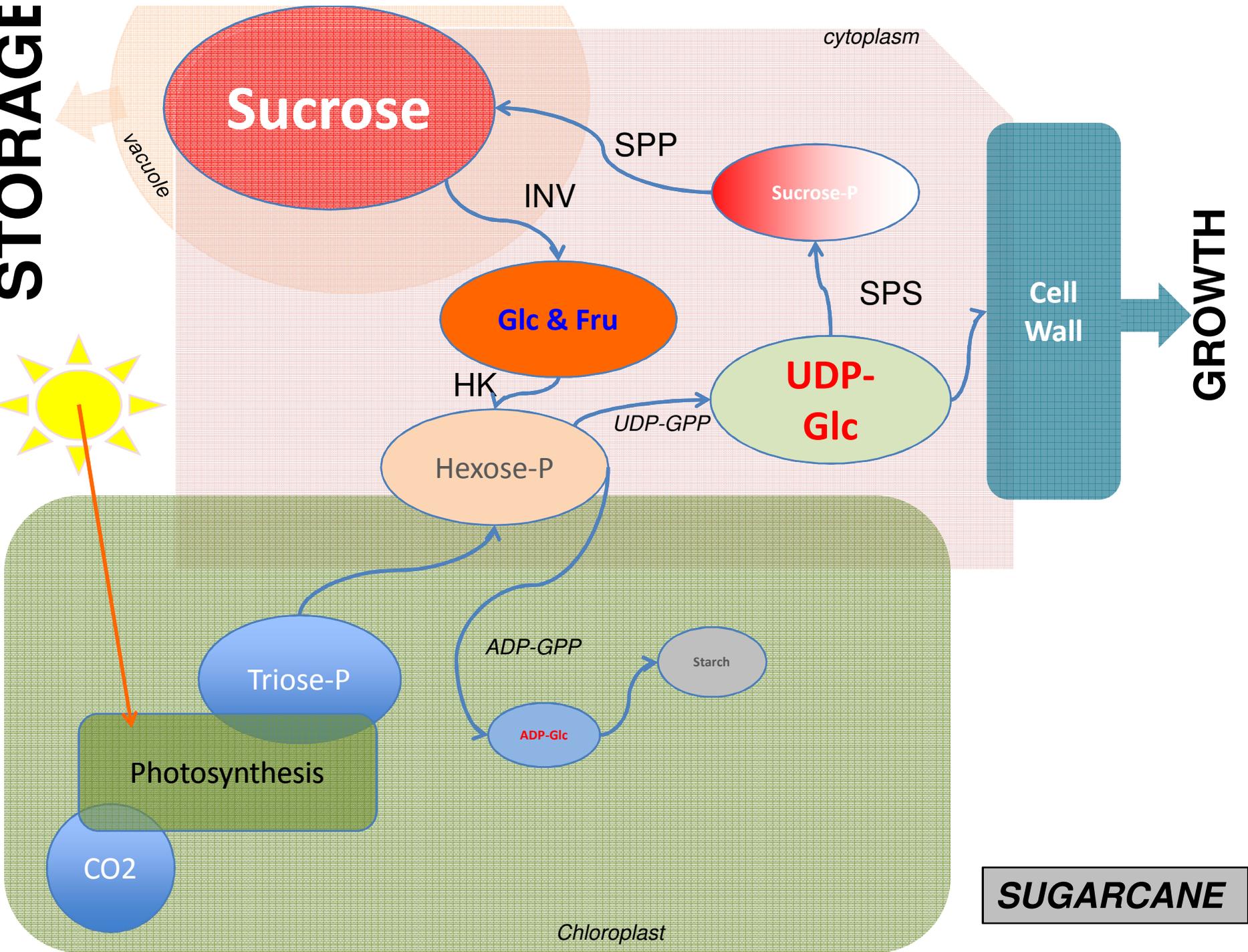
GROWTH

cytoplasm

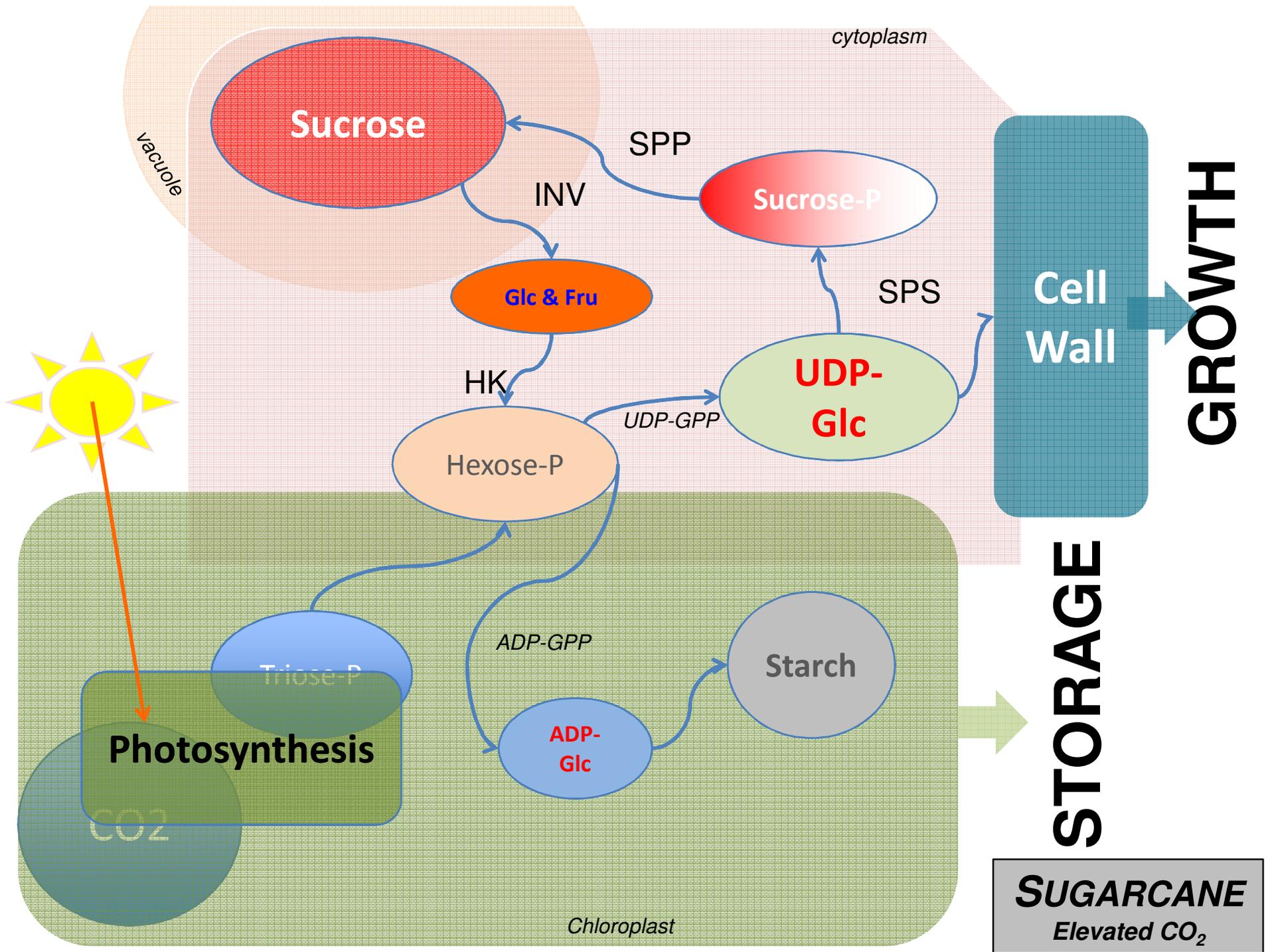
vacuole

Chloroplast

SUGARCANE

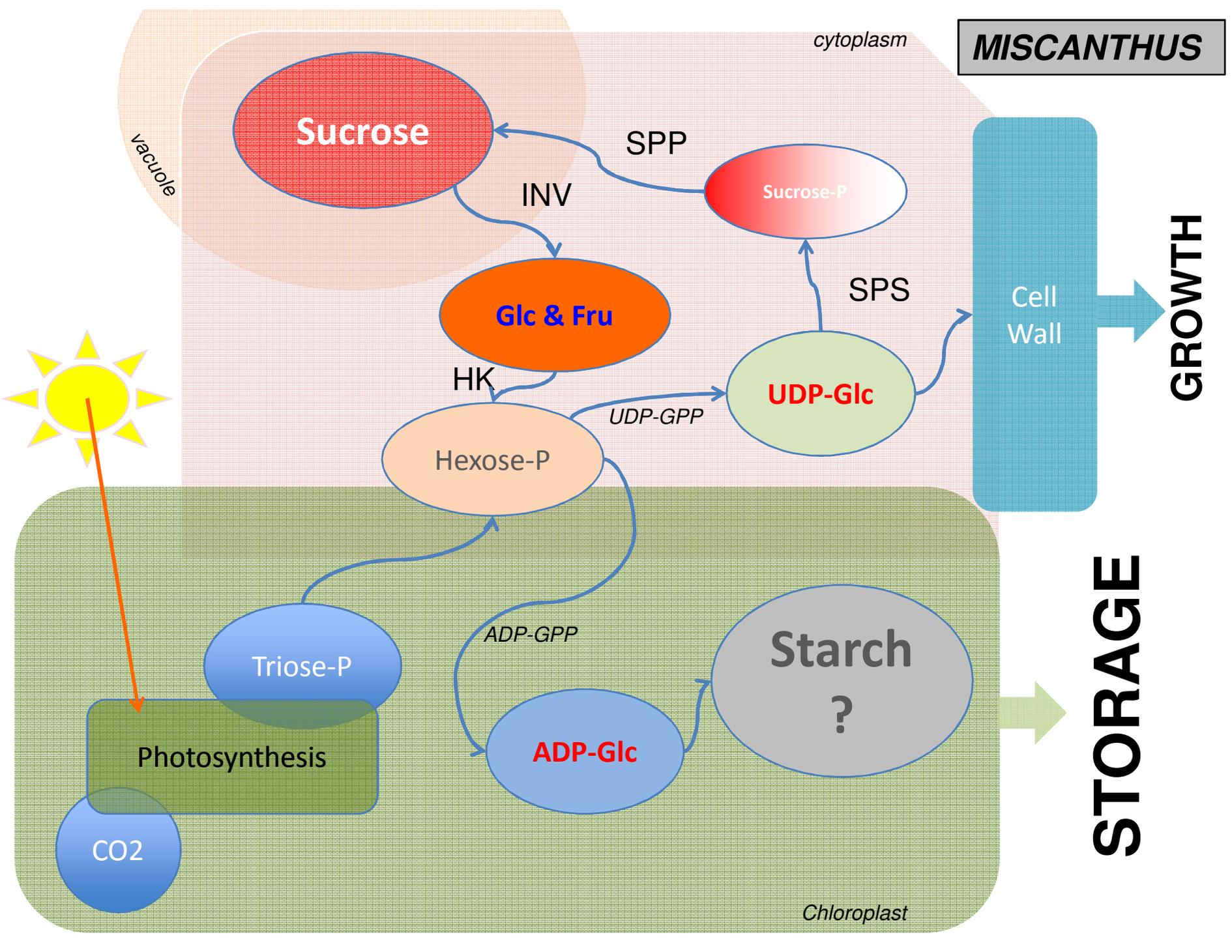


With elevated CO₂ sugarcane grows more and faster due to increase in ETR, but soluble sugars are lower and there is little starch accumulated in leaves. Instead of the ADP-glc pathway in the chloroplasts, sugarcane leaves seem to use the UDP-glc synthesis to foster cell wall biosynthesis and make more biomass



On the other hand, miscanthus does not accumulate sucrose as a C storage, as does sugarcane. Thus, it probably uses the UDP-glc pathway to synthesise cell walls and grow but possibly uses the ADP-glc pathway with the same intensity and produces starch in its leaves.

MISCANTHUS

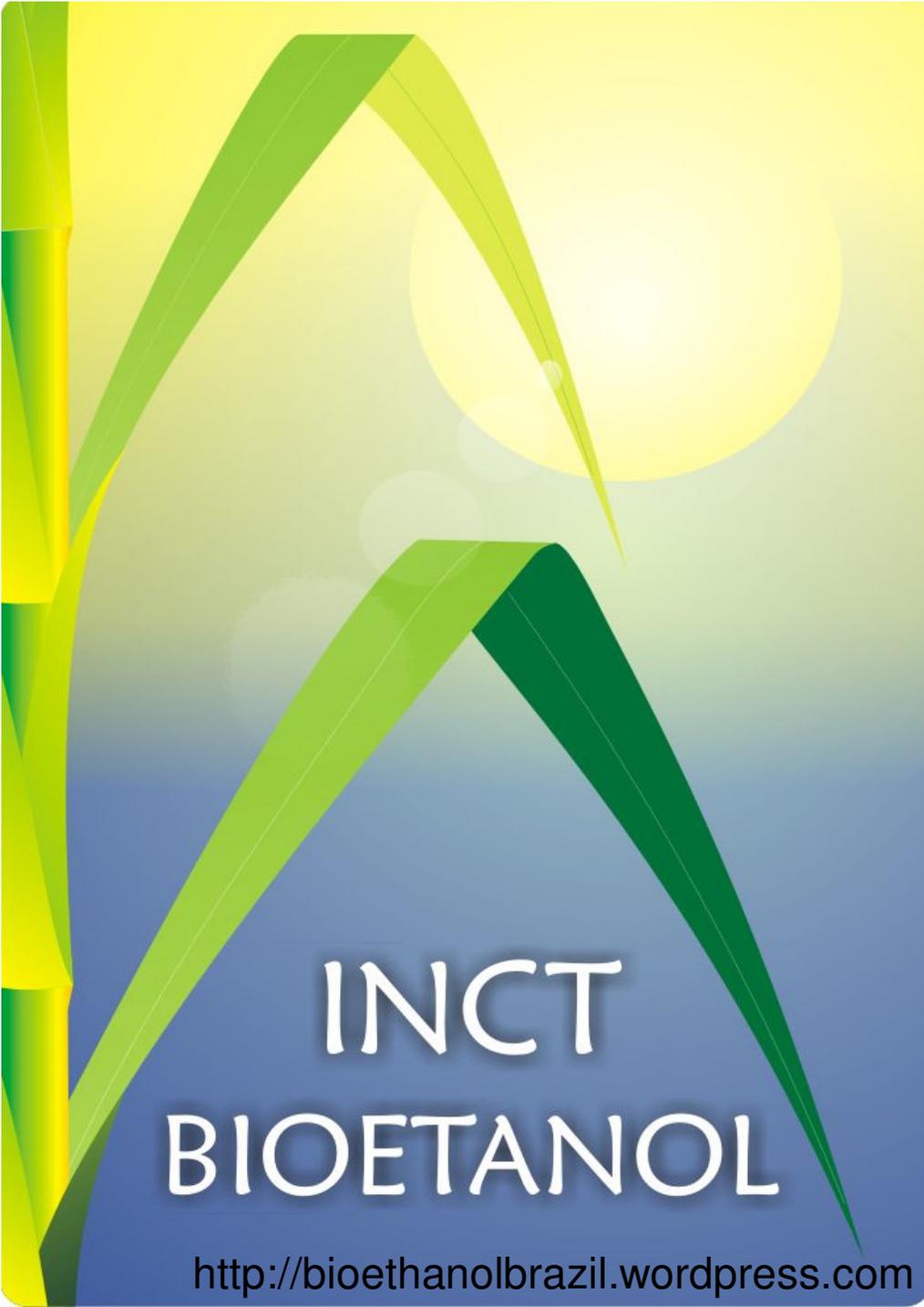


PERSPECTIVES FOR BIOFUELS

A comparison miscanthus X sugarcane would afford to find **network nodes** related with the accumulation of sucrose or cell walls in both species.

This might open the way to **genetically transform each species** so that they could be used either for bioethanol production from sucrose or cellulosic bioethanol

producing starch in sugarcane can be useful as in this way, **more carbon can be “packed”** than when sucrose is stored in vacuoles.



INCT BIOETANOL

<http://bioethanolbrazil.wordpress.com>

THANK YOU

msbuck@usp.br

